

Acute and Chronic Toxicity Studies of Ethanol Leaf Extract of *Merremia Tridentata* (Linn) Hallier F.

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ABSTRACT

The acute and chronic toxicity evaluation of Ethanol leaf extract of *Merremia tridentata* (Linn) Halier F. (MTELE) was carried out on albino wistar rats. Phytochemical screening and acute toxicity profile of the extract were determined using standard methods. The animals were assigned into groups and administered varying doses of MTELE (100, 200, 400 mg/kg body weight and 0.2 ml of distilled water) for a period of hundred days (fourteen weeks). The body weight, relative organ weight, haematology, serum biochemical indices and histopathological studies of the liver, kidney, spleen, heart, and lungs were appropriately carried out to determine propensity of possible toxicity. Phytochemical screening revealed the presence of alkaloids, tannins, cardiac glycosides, saponins, steroids, triterpenes, flavonoids while anthraquinone and cyanogenic glycosides were absent. The median lethal dose LD₅₀ was estimated as 2200 mg/kg body weight. There was significant ($p < 0.05$) reduction in the percentage change in body weight of rats administered 200 and 400 mg/kg/day dose of the extract for 100 days when compared to the control group. Moreover, there was a significant ($p < 0.05$) reduction in the relative weight of the spleen of rats and significant ($p < 0.05$) increase in the relative weight of the liver, kidney, heart and lungs of rats administered 400 mg/kg/day dose. All serum biochemical parameters studied showed significant ($p < 0.05$) increase in group administered 400 mg/kg body weight dose while alkaline phosphatase, aspartate amino transferase, creatine kinase, lactate dehydrogenase and potassium ion showed significant increase ($p < 0.05$) in the group administered 200 mg/kg/day. There is no significant change in hematological parameters like RBC, hemoglobin, hematocrit, platelets, monocytes, basophils, MCV, MCH, MCHC, in the extract treated animals except the lymphocyte that showed a significant ($p < 0.05$) reduction only in the group treated with 400 mg/kg body weight dose. Administration of MTELE at 200 mg/kg body weight did not occasioned any histo-architectural change in the liver and spleen but caused varying degree of remarkable histological derangement in the other tissues. Furthermore, there were remarkable pathologies in the liver, kidney, spleen, heart and lungs ranging from vascular congestion, haemorrhage, fibrosis, to renal and myocardial damage in the group treated with 400 mg/kg/day dose for hundred days. However, 100 mg/kg body weight dose showed no significant difference ($p > 0.05$) in all the parameters evaluated indicating safety at this dosage. Ethanol leaf extract of *Merremia tridentata* (Linn) Halier F. (MTELE) may not be safe at chronic administration even at dosage as low as 200 mg/kg body weight. The plant should be cautiously employed to avoid unwarranted complication on long term administration.

Keywords: Acute toxicity, Chronic toxicity, Evaluation, *Merremia tridentata* (Linn) Halier F.

I. INTRODUCTION

Medicinal plants have come to play important role as therapeutic alternative in many parts of the world. This paradigm shift has opened new frontier in global healthcare delivery with companies making fortune from these plants based on their folklorish use by herbal practitioners through the ages. *Merremia tridentata* (Linn) Halier F. family (*Convolvulacea*) is a plant found in wasteland as perennial shrub. It is a tropical climbing herb with thick rootstock spreading on walls and fences. In Nigeria, the plant is found

mainly in the North eastern and North western regions where it is employed in the treatment of scorpion sting, diabetes, and sugar related diseases, urinary and known locally as *Yambururu* in Hausa language [1]. The plant has been reported to be employed in traditional systems of medicine for treatment of various ailments such as piles, swellings, ulcers, rheumatic affections, stiffness of the joints, hemiplegia, urinary infections, and general debility [2], [3]. Reference [4] reported the anti-inflammatory capacity of the root extract of the plant. Analgesic and anti-inflammatory activities of these extracts were assessed in rats with hot plate

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test, writhing test in mice, carrageenan induced paw oedema and histamine induced paw edema in rats. Also, study on the anti-diabetic effect of the aqueous root extract of *M. tridentata* had been carried out [5] and the aerial parts were reported to contain flavonoids, diosmetin, luteolin, and their 7-O- β -D-glucosides [2] imparting on its antioxidant and free radical scavenging activities [3]. Other previous studies stated that *M. tridentata* have strong wound healing, anti-inflammatory, and anti-arthritis activities [6], [7]. Reference [8] reported that *M. tridentata* was used as a supplementary feed to the grass *Panicum maximum* for young West African Dwarf Sheep. However, in our literature search, there was no record on the safety of the plant especially on the long term hence the present study is aimed at investigating the acute and chronic toxicity profile of *Merremia tridentata* ethanol leaf extract (MTELE).

II. MATERIALS AND METHOD

A. Plant Material and Extraction

The aerial part of *Merremia tridentata* was collected from the bank of Tamburawa river along Zaria road Kano Nigeria. and was authenticated at the Herbarium unit of Botany department of Ahmadu Bello University Zaria, Nigeria. The plant material was shade dried for three weeks to allow the leaves to drop from the stem and pulverized using a kitchen blender. The powdered leaf sample was cold macerated using 70% ethanol for seventy two hours after which the solvent was evaporated using Rotary Vacuum Evaporator (Hahn Vapor, HS-2005V, Hahnshin Scientific Co., Korea) to obtain the *Merremia tridentata* ethanol leaf extract (MTELE) used for this study.

B. Animals

Healthy adult albino male rats weighing 100 to 150 g were obtained from National Animal Production and Research Institute (NAPRI, Vom, Nigeria) were grouped and housed in cages under 12 hours light dark cycle at 34 \pm 2 °C with free access to standard pellet diet and water *ad libitum*. The animals were allowed to acclimatize to the laboratory conditions for two weeks prior to the commencement of the experiment.

C. Chemicals

All chemicals and reagents used in this study are of analytical grade. Kits for estimation of biochemical parameters (Protein, Albumin, Creatinine, ALT, AST, ALP, LDH, Creatine kinase) are product of RANDOX Laboratory Ltd. Ardmores United Kingdom.

D. Acute Toxicity Study

The toxicity test was carried out on albino rats (100–120 g) in accordance with the protocol of Reference [9]. On the first phase, animals were grouped into three of three rats per group and administered 10, 100 and 1000 mg/kg body weight doses respectively of MTELE orally and monitored for twenty four hours for signs of toxicity like, tremor, restlessness, lacrimation, dizziness and death. In the second phase, another set of rats (one per group) were given oral dose of MTELE at 1200, 1600, 2900 and 5000 mg/kg respectively and monitored for twenty four hours for signs of toxicity. The

Median lethal dose (LD₅₀) was estimated from the Geometric Mean of the Maximum dose survived by all the rats and the minimum dose that caused fatality in all rats in the experiment [9].

E. Experimental Design for Chronic Toxicity Study

Albino male rats weighing 120 \pm 20 g was divided into four cages of five rats in each cage. They were placed on rat chow and allowed to drink water *ad libitum*. Group I was administered distilled water as the control while the remaining groups were administered specified doses of MTELE orally once a day for a period of fourteen weeks.

The treatments were as follows:

Group I: Normal rats treated with 0.2 ml distilled water.

Group II: Normal rats administered with 100 mg/kg body weight MTELE

Group III: Normal rats administered with 200 mg/kg body weight MTELE

Group IV: Normal rats administered with 400 mg/kg body weight MTELE

F. Body Weight, Relative Organ Weight and Feed Intake

The body weights of the animals in each group were recorded on weekly basis while the water and feed intake were recorded on daily basis all through the period of the experiment. On the 101st day, at the end of the experimental period for chronic studies, animals in all the groups were weighed and sacrificed by cervical decapitation after anaesthetizing them with Chloroform. Blood was collected in EDTA tube and plain tubes for the analysis of hematological and biochemical parameters respectively [10]. The sacrificed animals were dissected to harvest organs such as the liver, spleen, kidney, heart, and the lungs. These organs were washed with ice cold saline blotted dry and weighed.

G. Haematology and Biochemical Analysis

Blood samples collected on the day after the expiration of 100day administration of MTELE into EDTA container for hematological estimation were analyzed using auto Hematology Analyzer (Mindray, BC-2800) while blood for biochemical estimations collected in plain specimen bottles was centrifuged at 3000 rpm for 10 minutes to obtain the serum for analysis. Serum total protein, albumin, alanine amino transferase (ALT), aspartate amino transferase (AST), bilirubin (total and direct), alkaline phosphatase, urea, uric acid, creatinine, creatine kinase and lactate dehydrogenase levels were measured using biochemical assay kits (RANDOX Laboratory Ltd. Ardmores United Kingdom). Serum electrolytes (Sodium and Potassium ions) were estimated using the method described by [11].

H. Histopathological Examination

The liver, kidney, heart, lungs, and spleen were removed and fixed in 10% formal saline. The fixed tissues were transferred to a graded series of ethanol and then cleared in xylene. Once cleared, the tissues were infiltrated in molten paraffin wax in the oven at 58 °C. Serial section of 5 micrometers thick were obtained from a solid block of tissue, cleared, fixed in clean slides, stained with Haematoxylin and Eosin stains, and examined with the light microscope [12].

I. Statistical Analysis

Results were expressed as mean \pm SD. All data was subjected to one-way analysis of variance (ANOVA) and Tukey's post hoc test at 95% level of significance using MINITAB 17 statistical software.

III. RESULTS

A. Phytochemical Screening

The extract was found to contain the following secondary metabolites: Alkaloids, tannins, cardiac glycosides, saponins, steroids, triterpenes, flavonoids while anthraquinone and cyanogenic glycosides are absent (Table I).

B. Acute Toxicity Test

The result of the acute toxicity test is shown in Table II. In the first phase of the experiment, there was no mortality of any of the rats at 10, 100 and 1000 mg/kg body weight dose while mortality was only recorded at 5000mg/kg body weight dose. The median lethal dose was estimated at 2200 mg/kg body weight.

C. Effect of Different Doses on Body Weight Upon Chronic Administration

The change in weight of rat administered 100 mg/kg, 200 mg/kg and 400 mg/kg of MTELE is shown in Figure 1. There was no significant ($p>0.05$) difference in the mean body weight of the different treatment groups treated with the leaf extract of *M. tridentata* until after twelve weeks of treatment. Significant ($p<0.05$) difference in the weight gained in rats especially between the group treated with 400 mg/kg body weight dose and the control group from the thirteenth week was observed. The group administered with 200 mg/kg body weight of MTELE only had a significant ($p<0.05$) difference in weight gained at the fourteenth week.

D. Effect of the Different Doses of MTELE on Relative Organ Weight upon Chronic Administration

Administration of MTELE at two dosage levels tested (100 and 200 mg/kg body weight) did not caused any notable difference in the relative weight of the liver, kidney, lung, and heart (Table III). However, there was a significant ($p<0.05$) reduction in the relative weight of the spleen of rats administered 400 mg/kg body weight dose (Group IV) and significant increase ($p<0.05$) in the relative weight of the liver, kidney, heart, and lungs of rat administered 400mg/kg/day doses of MTELE for a period of fourteen weeks.

A. Effect of the Different Doses of MTELE on Haematology and Serum Biochemical Parameter upon Chronic Administration

The extract did not cause any alteration in the haematological parameters at all the dosages administered whereas the extract at 400 mg/kg body weight dose caused a significant ($p<0.05$) alteration in all the biochemical parameters investigated in this study. Furthermore, there was also significant increase in the serum level of potassium ion and activity of alkaline phosphatase, aspartate amino transferase, creatine kinase and lactate dehydrogenase in the group administered 200 mg/kg body weight of MTELE when

compared with the control group after hundred days of administration (Table VI).

On the biomarkers of kidney function, the ethanolic leaf extract caused a significant ($p<0.05$) increase in the blood urea, *creatinine and Potassium* at 200 mg/kg and 400 mg/kg body weight. The extract also caused a significant ($p<0.05$) reduction in serum concentration of sodium at 400 mg/kg body weight. Furthermore, the extract elicited a significant ($p<0.05$) increase in the level of creatine kinase and lactate dehydrogenase in the serum of the rat groups administered 200 and 400 mg/kg body weight doses of MTELE. The extract however did not show any effect different from the control group upon administration at 100mg/kg dose level.

B. Effect of the Different Doses of MTELE on Histo-architecture of Liver, Kidney, Heart, Spleen and Lungs upon Chronic Administration

Liver, kidney, heart, lung, and spleen of rat treated with 100mg/kg body weight dose of extract showed no remarkable pathology when compared to the control group. However, the liver from the group of rats administered 200 mg/kg body weight dose of extract for hundred days (fourteen weeks) showed unremarkable histo-architecture (Plate 1c) while the liver architecture from the 400 mg/kg body weight dose group showed a vast area of vascular congestion (Plate 1d). Moreover, the kidney of rat administered 200 mg/kg body weight dose of extract showed areas of haemorrhage and vascular congestion (Plate 2c) while the architecture of kidney of rat administered 400 mg/kg body weight dose for a period of 100 days revealed haemorrhage, vascular congestion, and renal damage (Plate 2d). There was no damage done to the spleen by the administration of MTELE at the different dose levels until 400 mg/day where fibrosis was conspicuously noticed (Plate 3d). The heart, which is another important organ in the living system reacted to the administration of MTELE by developing haemorrhage and vascular congestion at 200 mg/kg body weight dose and haemorrhage with myocardial damage at 400 mg/kg/day dose level. The Photomicrographs of the lung of rats administered 200 mg/kg and 400 mg/kg dose of *tridentata* leaf extract however showed areas of haemorrhage and vascular congestion (Plate 5c and d).

IV. DISCUSSION

Herbal medicine is well accepted as alternate source of treatment globally with the major constraint of the knowledge of the safe therapeutic dose especially in treatment involving a considerable length of time. The absence of scientific validation and toxicity evaluation had really called for concern in the use of plant extracts for quality health care delivery. Thence it is necessary to carry out toxicity evaluation of these plants with the intention to ascertain their safety at short and long term. It is the result of the toxicity testing using modern scientific medium that determines the acceptability and/or safety of use or otherwise of the extract. Toxicological studies had been carried out on a number of natural extracts [13]-[17]. In this study, the median lethal dose of MTELE (2200 mg/kg body weight) signifies that the extract is not entirely safe upon administration; low toxicity by interpretation [18]. The extract is made up of alkaloids,

carbohydrate, cardiac glycosides, flavonoids, saponins, steroids, tannins, and triterpenes. These phyto-constituents are known to possess an array of therapeutic properties ranging from antimicrobial, antiviral, antihelminthic, antifungi, antidiabetic, immunostimulatory, antidepressant, antitumour, lipemic, anti-inflammatory to antisickling [19], [1], [10]. Some of these secondary metabolites have been reported to be toxic in some experimental studies [20]. Reduction in body weight and internal organ weight are considered as sensitive indices of toxicity after exposure to toxic substances [21], [22]. There was a significant ($p < 0.05$) reduction in the percentage change in weight of rats administered 200 and 400 mg/kg body weight dose of MTELE when compared with the control group. This may be as a result of deficit in nutrient utilization and bioavailability for a period thereby resulting in growth retardation of the experimental animals. This may be a pointer the possibility of toxicity of the extract to these animals on chronic exposure.

There was also a significant ($p < 0.05$) reduction in the relative organ weight of the spleen of rats administered 400 mg/kg/day, and increase in the relative weight of the liver, kidney, heart and lungs in rat treated with 400 mg/kg/day of MTELE for 100 days. The spleen is a blood-forming organ in early life and later a storage organ for red corpuscles and platelets; because of the large number of macrophages, it also acts as a blood filter, both identifying and destroying effete erythrocytes. The atrophy of the spleen as seen in the effect of 400 mg/kg dose of MTELE could be taken as the response to the noxious stimuli exerted by the dose causing serious challenge to the enormous functions of the spleen. The liver is an essential organ responsible for detoxification among other functions alongside the kidney which helps in the excretion of such unwanted substances from the body system. The lung is also very essential in the process of respiration for energy generation to sustain life and wholesome development. The discrepancies brought about by this extract will definitely affect these vital functions and consequently the well being of the animals. Organ weight analysis is an important endpoint for identification of potentially harmful effects of test compounds in toxicology studies [23]. Moreover, organ weight is one of the most sensitive indicators of an effect of test article, as significant differences in organ weight between treated and untreated (control) animals may occur in the absence of any morphological changes [24]. Hence in this study, the effect of the extract on the liver, heart, lungs, and spleen may be a pointer to its potential toxicity but needs further confirmation using other indicators.

There is no significant ($p > 0.05$) change in hematological parameters like RBC, hemoglobin, hematocrit, platelets, monocytes, basophils, MCV, MCH, MCHC, in the extract treated animals. Hematological change such as anemia is reported to be accompanied with bone marrow toxicity [25], [26]. The white blood cells did not also show any significant ($p > 0.05$) difference in all the treated groups when compared with the control. It was only the lymphocyte that showed a significant ($p < 0.05$) reduction only in the group treated with 400 mg/kg body weight dose. This may be as a result of the atrophy of the spleen at this dose level indicating the possibility of toxicity to immune-stimulatory functions. The observed values of blood parameters within the normal range

suggest that the drug may be non-toxic in nature to the erythropoietic system at 100, 200 and 400 mg/kg/day.

Chronic exposure of the animals to the plant extract *Merremia tridentata* at 100 mg/kg dosage when tested for the biochemical parameters such as protein, urea, AST, ALT, LDH and creatinine do not show any significant difference in their levels when compared with the control animals. However, at higher dosage of 200 mg/kg/day for 100 days there were significant increase in the serum levels of alkaline phosphatase, aspartate amino transferase, lactate dehydrogenase and potassium ion while the group treated with 400 mg/kg/day manifested significant difference in serum levels of all the biochemical parameters investigated when compared with the control. The abnormal values of the biochemical parameters such as Urea and Creatinine suggest that the extract caused serious disturbance in the renal function. The transaminases (alanine amino transferase and aspartate amino transferase), creatine kinase, LDH, and alkaline phosphatases are good indices of liver, heart, and kidney damage respectively [27]-[30], [10]. Hence, the raised levels of these serum biochemicals are indication of potential toxicity of the extract to the liver, heart, and the kidney. The chronic administration of the leaf extracts of *Merremia tridentata* (Linn) elicited responses in some serum electrolyte concentration.

The extract administered in this chronic toxicity study led to significant ($p < 0.05$) increase in serum level of Potassium ion at 200 mg/kg and 400 mg/kg respectively while it caused a significant reduction ($p < 0.05$) in serum level of Sodium ion (Na^+) at 400 mg/kg dose. Hyperkalaemia is reported to occur due to severe renal injury, ketoacidosis or insulin deficiency and low levels of sodium ions reported to be caused by salt-losing nephritis, cirrhosis, gastro intestinal fluid loss and congestive heart failure [31]. The effect of these extracts at the higher doses tested on these electrolyte profile may be a pointer that the extract may be toxic to the process of electrolyte balancing in the kidney or to the functional unit of the kidney, liver, and the heart.

Histopathological studies of the liver, spleen, the kidneys, lungs, and the heart were carried out to confirm if the extract shows any morphological sign of toxicity on histo-architecture of any of these organs. MTELE at the 200 mg/kg dose did not cause any remarkable pathological architecture of the spleen and liver but resulted to haemorrhage and vascular congestion in the kidney, heart, and the lungs. The higher dose, 400 mg/kg of MTELE caused renal, myocardial damage, fibrosis and vascular congestion in the liver, spleen and lungs. This finding confirmed the result of the biochemical assay of the indices of organ damage which pointed at the extract at 200 mg/kg and 400 mg/kg doses as a potential culprit. Fibrosis is the formation of excess connective tissue in a reparative or a reactive process. The persistent stimulus of chronic inflammation activates macrophages and lymphocytes leading to the production of growth factors and cytokine which increase the synthesis of collagen. This usually alters the architecture and affects the functions of the organ concerned. In any of these pathological situations occasioned by chronic administration of MTELE at 200 and 400 mg/kg doses, the integrity of the affected organs was compromised. This indicates that the plant extract is not harmful at 100 mg/kg body weight dosage level and can be

safely used at dosage level much lower than 200 mg/kg body weight.

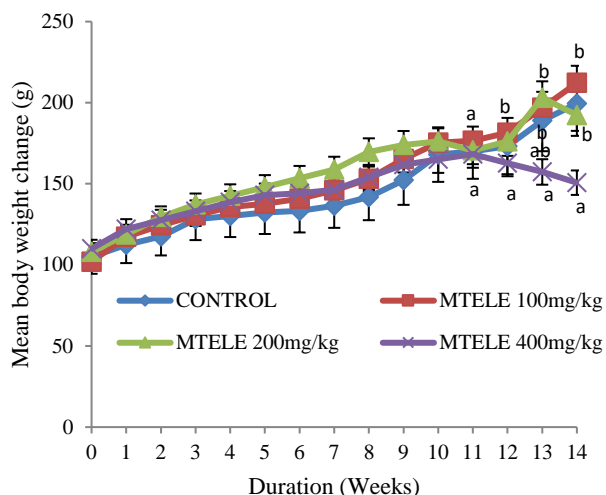


Fig 1: Effect of Administration of Ethanol Leaf Extract of *M. tridentata* on Mean Body Weight Change of Rats Groups for Fourteen Weeks.

^{a-c} Values with different letters near the lines for a given week are significantly ($p < 0.05$) different from each other group.

TABLE I: PHYTOCHEMICAL CONSTITUENT OF ETHANOL LEAF EXTRACT OF

MERREMIA TRIDENTATA	
Phytochemical	Status
Alkaloid	+
Anthraquinone	-
Glycosides	+
Cardiac Glycosides	+
Cyanogenic Glycoside	-
Flavonoids	+
Saponins	+
Steroids	+
Tannins	+
Triterpenes	+

+ Indicates presence and - indicates absence.

TABLE II: ACUTE TOXICITY STUDY OF MTELE

Dose (mg/kg body weight)	Mortality Index*	LD ₅₀
Phase 1: 10	0/3	
100	0/3	
1000	0/3	
Phase 2: 1200	0/1	
1600	0/1	
2900	1/1	
5000	1/1	$\sqrt{1600 \times 2900}$ = 2200

* Number of dead animal in the group
Total number of animals in the group

TABLE III: EFFECT OF ETHANOL LEAF AND STEM EXTRACTS OF *M. TRIDENTATA* ON RELATIVE ORGAN WEIGHT IN THE CHRONIC TOXICITY STUDY IN RATS

Treatment	Spleen	Liver	Kidneys (g/100 g)	Lung	Heart
Normal Control	0.30 ± 0.05 ^a	3.36 ± 0.30 ^a	0.43 ± 0.05 ^a	0.79 ± 0.17 ^a	0.42 ± 0.05 ^a
Normal Rat + MTELE (100 mg/kg)	0.29 ± 0.04 ^a	3.24 ± 0.26 ^a	0.42 ± 0.04 ^a	0.78 ± 0.16 ^a	0.51 ± 0.12 ^a
Normal Rat + MTELE (200 mg/kg)	0.39 ± 0.09 ^a	3.68 ± 0.05 ^a	0.50 ± 0.08 ^a	1.02 ± 0.19 ^a	0.62 ± 0.16 ^{ab}
Normal Rat + MTELE (400 mg/kg)	0.17 ± 0.08 ^b	4.00 ± 0.11 ^b	0.93 ± 0.13 ^b	1.42 ± 0.29 ^b	0.99 ± 0.31 ^b

^{a-b} Values expressed as Mean ± SD of five animals with different letters along a column are significantly ($p < 0.05$) different from each other group of animals. MTELE means *M. tridentata* ethanol leaf extract.

TABLE IV: EFFECT OF ETHANOL LEAF EXTRACT OF *M. TRIDENTATA* ON HAEMATOLOGICAL PARAMETERS IN THE CHRONIC TOXICITY STUDY IN RATS

Parameters	Normal Control	Normal Rats + Extract (100 mg/kg)	Normal Rats + Extract (200 mg/kg)	Normal Rats + Extract (400 mg/kg)
Red blood cell ($\times 10^6/\mu\text{l}$)	4.87 ± 0.25 ^a	4.97 ± 0.25 ^a	4.07 ± 0.72 ^a	5.90 ± 0.26 ^a
Haemoglobin (g/dl)	13.50 ± 3.44 ^a	13.68 ± 2.75 ^a	13.87 ± 1.43 ^a	15.72 ± 1.56 ^a
Haematocrit (%)	40.91 ± 5.26 ^a	41.21 ± 6.88 ^a	42.03 ± 4.77 ^a	47.63 ± 4.48 ^a
M C V (fl)	8.40 ± 0.46 ^a	8.29 ± 0.44 ^a	10.32 ± 0.54 ^a	8.07 ± 0.41 ^a
M CH (pg)	27.72 ± 6.08 ^a	27.53 ± 4.76 ^a	34.07 ± 6.43 ^a	26.64 ± 10.69 ^a
M C H C (g/dl)	33.00 ± 0.00 ^a	33.07 ± 0.11 ^a	33.00 ± 0.00 ^a	33.00 ± 0.15 ^a
Platelet ($\times 10^5/\mu\text{l}$)	139.33 ± 48.06 ^a	116.67 ± 15.27 ^a	130.67 ± 5.03 ^a	120.00 ± 20.00 ^a
White blood cell ($\times 10^3/\mu\text{l}$)	3.13 ± 1.55 ^a	4.02 ± 0.31 ^a	4.42 ± 0.45 ^a	4.44 ± 0.20 ^a
Neutrophil (%)	54.65 ± 0.58 ^a	54.33 ± 3.12 ^a	53.33 ± 3.57 ^a	52.33 ± 2.83 ^a
Lymphocyte (%)	42.63 ± 0.57 ^a	42.68 ± 1.34 ^a	43.32 ± 2.55 ^a	46.00 ± 3.90 ^a
Monocytes (%)	1.67 ± 0.58 ^a	2.00 ± 0.62 ^a	2.33 ± 0.53 ^a	1.00 ± 0.00 ^a
Eosinophil (%)	1.00 ± 0.00 ^a	0.67 ± 0.22 ^a	1.00 ± 0.00 ^a	0.67 ± 0.18 ^a
Basophil (%)	0.00 ± 0.00 ^a	0.00 ± 0.00 ^a	0.00 ± 0.00 ^a	0.00 ± 0.00 ^a

^{a-b} Values expressed as Mean ± SD of five animals. Values with different letters along a row are significantly ($p < 0.05$) different from each other. MCV- Mean Corpuscular volume; MCH- Mean Corpuscular Haemoglobin; MCHC- Mean Corpuscular Haemoglobin Concentration.

TABLE V: EFFECT OF ETHANOL LEAF EXTRACT OF *M. TRIDENTATA* ON BLOOD BIOCHEMICAL PARAMETERS IN THE CHRONIC TOXICITY STUDY IN RATS

Parameters	Normal Control	Normal Rats + Extract (100 mg/kg)	Normal Rats + Extract (200 mg/kg)	Normal Rats + Extract (400 mg/kg)
Total Protein (g/L)	65.33 ± 3.44 ^a	63.91 ± 3.86 ^a	65.74 ± 4.15 ^a	89.38 ± 4.73 ^b
Albumin (g/L)	36.99 ± 3.11 ^a	37.26 ± 2.74 ^a	35.91 ± 1.23 ^a	47.34 ± 1.32 ^b
Albumin/Globulin	1.31 ± 0.12 ^a	1.44 ± 0.29 ^a	1.22 ± 0.25 ^a	1.13 ± 0.06 ^b
Total Bilirubin (mmol/L)	2.74 ± 0.27 ^a	2.76 ± 0.26 ^a	2.78 ± 0.23 ^a	4.99 ± 0.24 ^b
Direct Bilirubin (mmol/L)	1.35 ± 0.11 ^a	1.52 ± 0.31 ^a	1.35 ± 0.08 ^a	2.27 ± 0.14 ^b
Urea (mmol/L)	5.91 ± 0.13 ^a	5.89 ± 0.13 ^a	6.29 ± 0.48 ^a	11.15 ± 2.00 ^b
Creatinine (mmol/L)	51.56 ± 2.34 ^a	51.08 ± 2.09 ^a	52.02 ± 3.86 ^a	74.33 ± 9.44 ^b
Alkaline Phosphatase (U/l)	61.28 ± 3.92 ^a	66.20 ± 4.67 ^{ab}	76.90 ± 8.59 ^b	116.52 ± 14.17 ^c
Alanine amino transferase (U/l)	33.40 ± 6.40 ^a	32.69 ± 4.73 ^a	35.66 ± 3.87 ^a	46.32 ± 6.81 ^b
Aspartate aminotransferase (U/l)	80.22 ± 2.49 ^a	77.63 ± 4.69 ^a	85.43 ± 5.21 ^b	84.49 ± 3.96 ^b
Creatine Kinase (IU)	0.61 ± 0.03 ^a	0.57 ± 0.05 ^a	0.68 ± 0.03 ^b	0.75 ± 0.04 ^c
Lactate Dehydrogenase (IU)	205.33 ± 4.45 ^a	201.31 ± 3.22 ^a	226.42 ± 8.08 ^b	240.67 ± 7.58 ^b
Sodium Ion (mmol/L)	140.20 ± 3.19 ^a	139.00 ± 1.92 ^a	139.80 ± 1.92 ^a	131.80 ± 6.30 ^b
Potassium Ion (mmol/L)	4.02 ± 0.31 ^a	4.06 ± 0.32 ^a	4.58 ± 0.16 ^b	5.30 ± 0.37 ^c

^{a-d} Values expressed as Mean ± SD of five animals. Values with different letters along a row are significantly ($p < 0.05$) different from each other.

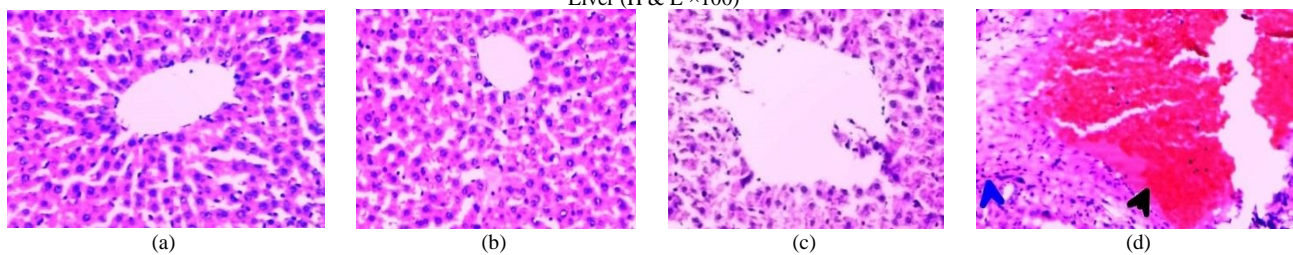
Liver (H & E $\times 100$)

Plate 1 a-d. (a) Photomicrograph of the liver of a normal control rat showing normal hepatocytes arranged in polygonal unit containing a central venule, (b) Liver of rats administered 100 mg/kg dose of *M. tridentata* ethanolic leaf extract showing intact tissue, (c) micrograph showing no damage (200 mg/kg), (d) Liver showing fibrosis and vast area of vascular congestion (400 mg/kg).

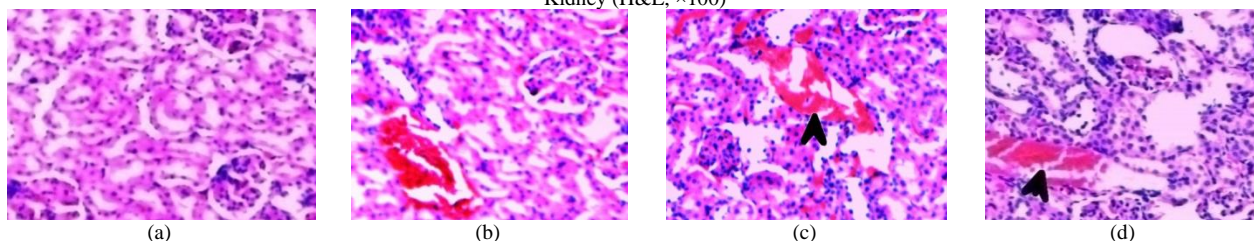
Kidney (H&E, $\times 100$)

Plate 2a-d: (a) Photomicrograph of the kidney of a normal control rat showing unremarkable kidney with the cortex containing glomeruli and medullary renal tubules, (b) kidney of rat administered 100 mg/kg MTELE showing intact tissue, (c): kidney showing haemorrhage and vascular congestion at 200 mg/kg dose, (d): kidney of rat administered 400 mg/kg MTELE showing areas of haemorrhage, vascular congestion (arrowed) and renal damage.

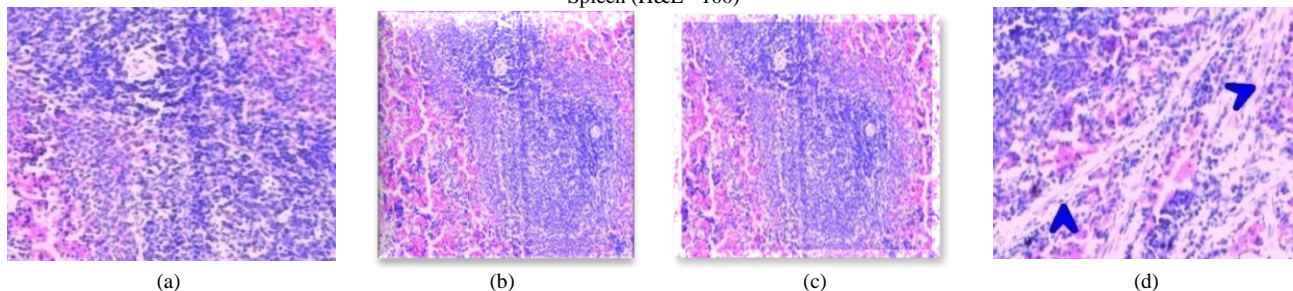
Spleen (H&E $\times 100$)

Plate 3a-d. (a) Photomicrograph of control rat spleen showing the red and white pulp (Normal architecture. b & c: spleen of rat treated with 100 mg/kg & 200 mg/kg dose MTELE respectively showing normal architecture. (d). spleen of rats treated with 400 mg/kg dose of MTELE showing areas of fibrosis (arrowed)

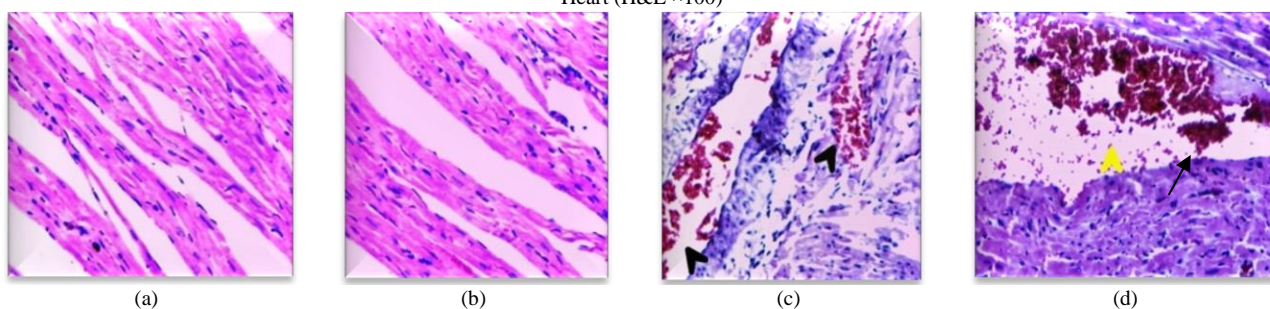
Heart (H&E $\times 100$)

Plate 4a-d. (a & b): Photomicrographs of the heart of a normal control rat and rat administered 100 mg/kg dose of MTELE showing an unremarkable myocardium. (c): heart of rat administered 200 mg/kg leaf extract showing haemorrhage and vascular congestion. (d): Photomicrograph of the heart of rat administered 400 mg/kg showing haemorrhage and myocardial damage.

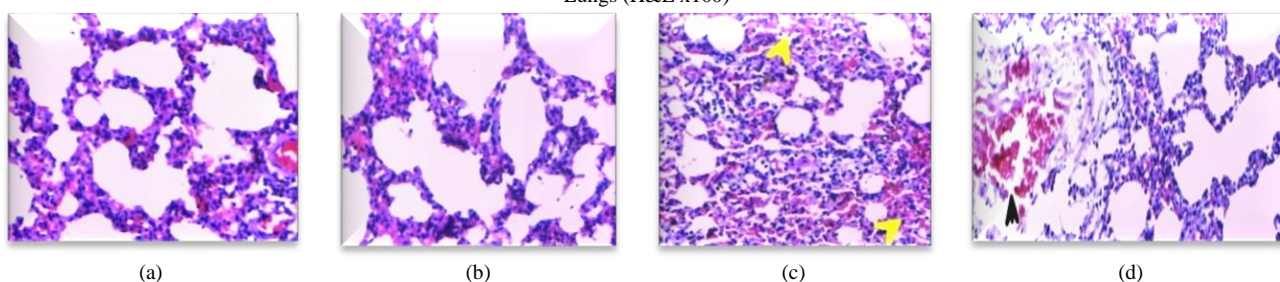
Lungs (H&E $\times 100$)

Plate 5a-d/ (a & b). Photomicrographs of the lung of a normal control rat and rat administered 100 mg/kg dose of MTELE showing normal architecture. (c & d): Photomicrographs of the lung of rats administered 200 mg/kg & 400 mg/kg dose of *tridentata* leaf extract showing areas of haemorrhage and vascular congestion.

V. CONCLUSION

Chronic administration of ethanol leaf extract of *Merremia tridentata* (Linn.) Hallier F. (MTELE) shows serious adverse effects on parameters such as body weight changes, relative organ weight and biochemical changes as indicated by the level of the different biomarkers of organ damage. Histopathological studies revealed different level of damage to the liver, kidney, spleen, heart, and lungs in the higher dose level tested in the study. It may be concluded that the ethanol leaf extract of *Merremia tridentata* (Linn.) Hallier F. (MTELE) may be considered relatively toxic at dosage level as low as 200 mg/kg body weight on chronic administration.

REFERENCES

- [1] M.O. Soladoye and O.O. Oyesiku, "Taxonomy of Nigerian Medicinal Plants" in *A Textbook of Medicinal Plants from Nigeria*, T. Odubgemi, Ed., Akoka-Yaba Lagos: Lagos University Press, 2008, ch 11, pp 93-149.
- [2] C.P. Khare, *Indian Medicinal Plants: An Illustrated Dictionary*. Springer, New Delhi, 2007, pp. 410-411.
- [3] Sowndhararajan, K and Nyuk, L.C. (2014). Antioxidant and Anti-ulcer Effects of Ethyl Acetate Fraction of *Merremia Tridentata* (L.) Hallier F. Root *Agriculture and Agricultural Science Procedia*, 2, pp. 406-414.
- [4] K. Arunachalam, T. Parimelazhagan and S. Manian, Analgesic and anti-inflammatory effects of *Merremia tridentata* (L.) hallier f. *Int J Pharm Pharm Sci* 2011; 3: pp75-79.
- [5] K., Arunachalam, and T. Parimelazhagan, Antidiabetic activity of aqueous root extract of *Merremia tridentata* (L.) Hall. f. in streptozotocin-induced-diabetic rats. *Asian Pacific Journal of tropical medicine*, 2012. 5(3), pp 175-179.
- [6] B.C. Hatapakki, V. Hukkeri, D.N. Patil, and M.J. Chavan. Wound Healing Activity of *Merremia Tridentata*. *Indian Drugs* 2004, 41, 532.
- [7] M., Kamalutheen, S., Gopalakrishnan, and T.S., Ismail. Anti-Inflammatory and Anti-Arthritic Activities of *Merremia Tridentata* (L.) Hall. f. *E. Journal of Chemistry*, 2009; 6, pp 943-948.
- [8] A., Aschfalk, H., Steingass, W., Muller and E. Drochner, *Merremia tridentata* as a Supplementary Feed to the Grass *Panicum Maximum* for Young West African Dwarf Sheep. *Tropical Animal Health and Production*, 2002, 34, pp 45-50.
- [9] D. Lorke, A new approach to practical acute toxicity testing. *Archives of Toxicology*, 1983, 54: pp 275 - 287.
- [10] P. Kalaiselvi, M., Uma, M., Suresh, K. Thulasiraman, and E. Lakshmidevi, Chronic toxicity studies of aqueous leaf extract of Indian traditional medicinal plant *Ocimum tenuiflorum* (Linn.) in rats *European Journal of Experimental Biology*, 2013, 3(5): pp 240-247.
- [11] Evenson M.E. (1999). "Spectrophotometric Technique". In: *Tietz Textbook of Clinical Chemistry*, 3rd ed, CA Burtis, ER Ashwood, Eds. Philadelphia:WB Saunders Co., pp. 75-93.
- [12] J. Ochei and A. Kolhatkar, "Histopathological Techniques and Cytology". In: *Textbook of Medical Laboratory Technology- Theory and Practice*, 3rd ed. 2008, Tata McGraw-Hill Publishing Company Limited, New Delhi: pp.418 - 426.
- [13] C.A. Pieme, V.N. Penlap, B. Nkegoum, C.L. Taziebou, E.M. Tekwe, F.X. Etoa and J. Ngongang, Evaluation of acute and subacute toxicities of aqueous ethanol extract of leaves of *Senna alata* (L.) Roxb (*Cesalpiniaceae*). *African Journal of Biotechnology*, 2006, 5(3): pp. 283-289.
- [14] K.C. Patrick-Iwuanyanwu, M.O. Wegwu and J.K. Okiyi Hepatoprotective effects of African locust bean (*Parkia clappertoniana*) and negro pepper (*Xylopi aethiopic a*) in CCl4 - induced liver damage in Wistar albino rats. *International Journal of Pharmaceutics* 6, 2010, pp.744-749.
- [15] S. Kumar, P.Singh, G. Mishra, S. Srivastar, KK. Jha, RL. Khosa, Phytopharmacological review of *Alternanthera brasiliana* (*Amaranthaceae*). *Asian Journal of Plants Science Research*, 2011.1(1): pp.41- 47.
- [16] C.U. Osifo, U. Akpamu, C.I. Idehen, W.A. Adisa, and K.E. Azeke, The effect of chronic ingestion of crude garcinia kola on the histology of the liver, *European Journal of Experimental Biology*, 2012. 2 (2): pp.404-409.
- [17] S.O. Ogbonnia, S, G.O. Mbaka, F.E. Nkemehule, J.E. Emordi, N.C. Okpagu and D.A. Ota, Acute and subchronic evaluation of aqueous extracts of *Newbouldia laevis* (Bignoniaceae) and *Nauclea latifolia* (Rubiaceae) roots used singly or in combination in Nigerian traditional medicines. *British Journal of Pharmacology and Toxicology*, 2014. 5(1): pp. 55-62.
- [18] W.J. Brock, H.J. Trochimowicz, R.J. Millischer, C. Farr, T. Kawano and G.M. Rusch, Acute and subchronic toxicity of 1,1-dichloro-1-fluoroethane (HCFC-141b). *Food and Chemical Toxicology*, 1995. 33, pp. 483-490.
- [19] A.J. Fridous, S. N. L. M. Islam and A.B.M. Faruque, Antimicrobial activity of the leaves of *Adhatoda vasica*, *Calotropis gigantea*, *Nerium odorum* and *Ocimum sanctum*. *Bangladesh. Journal of Botany*. 1990. 19: 227-229.
- [20] G.V. Pierangeli and L.R. Windell, Antimicrobial activity and cytotoxicity of *Chromolaena odorata* (L. f). King and Robinson and *Uncaria perrottetii* (A. Rich) Merr. extracts. *Journal Medicinal Plants Research*, 2009. 3(7): pp.511-518.
- [21] M. Raza, O.A. Al-Shabanah, T.M. El-hadiyah and A.A. Al-Majed Effect of Prolonged Vigabatrin Treatment on Hematological and Biochemical Parameters in Plasma, Liver and Kidney of Swiss Albino Mice. *Scientia Pharmaceutica*, 2002, 70, pp. 135-145.
- [22] S.K. Thanabhorn, S. Jenjoy, K. Thamaree, K. Ingkaninan and A. Panthong, Acute and subacute toxicity study of the ethanol extract from *Lonicera japonica* Thunb. *Journal of Ethnopharmacol.*, 2006. 107: pp. 370-373.
- [23] R. Nirogi, V.K. Goyal, S. Jana, S.K. Pandey, and A. Gothi, What suits best for organ weight analysis: review of relationship between organ weight and body / brain weight for rodent toxicity studies *International Journal of Pharmaceutical Science and Research*, 2014, Vol. 5(4): 1525-1532.
- [24] S.A Bailey, R.H. Zidell and R.W. Perry, Relationships between organ weight and body/brain weight in the rat: What is the best analytical endpoint? *Toxicol. Pathol.* 2004. 32, 448-466.
- [25] H. Rhiouani, J. El-Hilalya, Z.H. Israili and B. Lyoussia, Acute and Sub-chronic toxicity of an aqueous extract of the leaves of *Herniara glabra* in rodents. *Journal of Ethnopharmacology*, 2008, 118, 378-386.
- [26] R.K Koshiy, B.R Kapoor and M. Azmathulla, 2010. *Pharmacology online*, 3, pp. 229-242.
- [27] D.W. Martin P.A. Mayes and Y.M. Rodwell, In: Harper's Review of Biochemistry. 18th edn, Lange Medical, CA, 1981, pp:61.
- [28] R. Horton, L.A. Moran, R. Ochs, J.D. Rawn and K.G. Scrimgeour, *Principles of Biochemistry*. 2nd Edn., 1996, Prentice Hall.
- [29] K.M. Wasan, S. Najafi, J. Wong, and M. Kwong Assessing plasma lipid levels, body weight and hepatic and renal toxicity following chronic oral administration of a water soluble phytostanol compound. *Journal of Pharmaceutical Science*, 2001.4(3): 228-234.
- [30] M.A. Crook, *Clinical Chemistry and Metabolic Medicine*. 7th Edn., Hodder Arnold, London, 2006. pp. 426.
- [31] J. Ochei and A. Kolhatkar, *Textbook of Medical Laboratory Technology- Theory and Practice*, 3rd ed. 2008, Tata McGraw-Hill Publishing Company Limited, New Delhi: pp.180-182.